

PERFORMANCE CHARACTERISTICS

A) Internal Evaluation:

- Accuracy: In an internal study **Electra™ CA 15-3** was evaluated against commercially available licensed kit with 90 random clinical samples, & **Electra™ CA 15-3** has demonstrated 100% clinical correlation with the commercially available licensed kit.
- Precision: **Electra™ CA 15-3** was evaluated with licensed external Quality controls for Precision Studies & following is the data:

Controls	No. of testings	Mean Control values with Electra™ CA 15-3	Coefficient of Variation (CV)
Level 1	10	26.88	3.74
Level 2	10	52.30	2.61
Level 3	10	104.94	2.56

B) External Evaluation:

Electra™ CA 15-3 CLIA has been evaluated by a NABL accredited lab against their reference method. In this evaluation **Electra™ CA 15-3** has demonstrated 100% correlation with the reference method.

*Data file: Zephyr Biomedicals (A Division of Tulip Diagnostics Pvt. Ltd).

Important Note:

- The **Electra™ CA 15-3** assay is a temperature sensitive assay. The best temperature condition for this assay is from 18°C to 25°C.
- The wash procedure is critical. Insufficient washing will result in poor precision and falsely elevated absorbance readings.
- It is recommended to use the multiple channel pipettes to avoid time effect. A full plate of 96 wells may be used if automated pipette is available.
- Duplication of Standards & samples is not mandatory but may provide information on reproducibility & application errors.












LIMITATIONS OF THE ASSAY

(1). As with all diagnostic tests, a definite clinical diagnosis should not be based on the results of a single test, but should only be made by the physician after all clinical and laboratory findings have been evaluated. (2). The activity of the enzyme used is temperature-dependent and the RLU values may vary. The higher the room temperature (+18°C to +25°C) during substrate incubation, the greater will be the RLU values. Corresponding variations apply also to the incubation times. However, the Standards are subject to the same influences, with the result that such variations will be largely compensated in the calculation of the result. (3). Adaptation of this assay for use with automated sample processors and other liquid handling devices, in whole or in part, may yield differences in test results from those obtained using the manual procedure. It is the responsibility of each laboratory to validate that their automated procedure yields test results within acceptable limits. (4). Insufficient washing (e.g., less than 5 wash cycles, too small wash buffer volumes, or shortened reaction times) can lead to incorrect results.

BIBLIOGRAPHY

(1). Rongen HA, Hoetelmans RM, Bult A, van Bennekom WP. Chemiluminescence and immunoassays. **J Pharm Biomed Anal** 1994 Apr;12(4):433-62. (2). Koszegi T. Immunoluminometric detection of human prolactin. **J Biochem Biophys Methods** 2002 Oct;53(13):157-64. (3). Roda A, Simoni P, Mirasoli M, Baralini M, Violante FS. Development of a chemiluminescent enzyme immunoassay for urinary 1-hydroxypyrene. **Anal Bioanal Chem** 2002 Apr;372(7-8) (4). Laffin RJ, et al. Hybritech total and free prostate-specific antigen assays developed for the Beckman Coulter access automated chemiluminescent immunoassay system: a multicenter evaluation of analytical performance. (5). **Clin Chem** 2001 Jan;47(1):129-32. (6). Koszegi T, Walker W.H.C. Introduction: An Approach to Immunoassay. **Clin. Chem.** 1977; 23: 384. (7). Kirkegaard C., Friis T. and Siersback-Nielsen K. **Acta Endocrinol.** 1974; 77: 71. (8). Wisdom G.B. Enzyme-Immunoassay. **Clin. Chem.** 1976; 22: 1243. (9). Hoffenberg R. **Medicine** 1978; 8: 392. (10). Lieblisch J., Utiger R.D. **J. Clin. Invest.** 1972; 51: 1939.

SYMBOL KEYS

 Temperature Limitation	 Consult Instructions for use	 Date of Manufacture	 Batch Number / Lot Number
 Manufacturer	 In vitro Diagnostic Medical Device	 This side up	 Contains sufficient for <n> tests
 Use by	 Catalogue Number	 Do not reuse	

Manufactured by:

Zephyr Biomedicals

A Division of Tulip Diagnostics (P) Ltd.

M 46-47, Phase III B, Verna Industrial Estate, Verna, Goa- 403 722, INDIA.

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Certified Company

0422/VER-01

electra™
●●●●●●●●●● **CA 15-3**

Chemiluminescence Assay for the Quantitative Determination of Breast Cancer Antigen (CA 15-3) in Human Serum.

FOR IN VITRO DIAGNOSTIC USE ONLY
Store at 2°C to 8°C

INTENDED USE

ELECTRA™ CA 15-3 CLIA test is intended for the quantitative determination of Breast Cancer Antigen (CA 15-3) in human serum. For In Vitro Diagnostic Use only.

INTRODUCTION

Breast cancer is the most common life-threatening malignant lesion in women of many developed countries today, with approximately 180,000 new cases diagnosed every year. Roughly half of these newly diagnosed patients are node-negative, however 30% of these cases progress to metastatic disease. There are a number of tumor markers that can help clinicians to identify and diagnose which breast cancer patients will have aggressive disease and which will have an indolent course. These markers include estrogen and progesterone receptors, DNA ploidy and percent-S phase profile, epidermal growth factor receptor, HER-2/neu oncogene, p53 tumor suppressor gene, cathepsin D, proliferation markers and CA15-3. CA15-3 is most useful for monitoring patients post-operatively for recurrence, particularly metastatic diseases. 96% of patients with local and systemic recurrence have elevated CA15-3, which can be used to predict recurrence earlier than radiological and clinical criteria. A 25% increase in the serum CA15-3 is associated with progression of carcinoma. A 50% decrease in serum CA15-3 is associated with response to treatment. CA15-3 are more sensitive than CEA in early detection of breast cancer recurrence. In combination with CA-125, CA15-3 has been shown to be useful in early detection of relapse of ovarian cancer. CA15-3 levels are also increased in colon, lung and hepatic tumors.

PRINCIPLE

ELECTRA™ CA 15-3 Quantitative CLIA assay is for use on **ELECTRA™** analyzer. **ELECTRA™ CA 15-3 CLIA** works on the principle of chemiluminescence wherein light is produced by a chemical reaction from a substance as it returns from an electronically excited state to the ground state. When catalysed by HRP, the oxidation of luminol by hydrogen peroxide produces an electronically excited form of 3-aminophthalate which on relaxation emits light with maximum intensity at $\lambda=425\text{nm}$.

The CA 15-3 test is a solid phase two-site immunoassay. One monoclonal antibody is coated on the surface of the microtiter wells and another monoclonal antibody labeled with horseradish peroxidase is used as the tracer. The CA 15-3 molecules present in the standard solution or serum are sandwiched between the two antibodies, following the formation of the coated antibody-antigen-antibody-enzyme complex. The unbound antibody-enzyme labels are removed by washing and bound enzyme is detected by adding the chemiluminescent substrate. The bound enzyme converts substrate to a reaction product that emits a photon of light. Chemiluminescence is measured in Relative Light Units (RLU). The amount of light emitted is proportional to the amount of enzyme present and is directly related to the amount of CA 15-3 antigen in the sample. By reference to a series of CA 15-3 standards assayed in the same way, the concentration of CA 15-3 in the unknown sample is quantified.

MATERIALS & COMPONENTS

Materials provided with the test kits:

- Coated Microwells: Microwells coated with Anti- CA 15-3 antibody.
- Sample Diluent.
- CA 15-3 Enzyme Conjugate. Ready to use.
- Substrate A: Chemiluminescent substrate containing enhanced luminol solution.
- Substrate B: Chemiluminescent substrate containing stabilized peroxide solution.
- CA 15-3 Standard set of 6 standards labeled as A to F in liquid form. Ready to use. For standard Concentrations refer vial label.
- Wash Buffer Concentrate (20X).

Materials required but not provided:

- Precision pipettes: 10-100µl, 50-200µl, 100-1000µl
- Disposable pipette tips
- Distilled water
- Disposable Gloves
- ELECTRA™ Analyzer**

electra™

Chemiluminescence assay

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Chemiluminescence assay

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STORAGE AND STABILITY

1. **ELECTRA™ CA 15-3** kit is stable at 2-8°C up to the expiry date printed on the label.
2. Coated micro-wells should be used within one month of opening the pouch. Once opened, the pouch must be sealed properly to protect from moisture. In case the desiccant pouch changes color from blue to white, the strips should not be used.
3. Diluted wash buffer is stable up to one week at 2-8°C.
4. Working Substrate (A+B) must be used immediately.

SPECIMEN COLLECTION

1. Collect blood specimen by venipuncture according to the standard procedure.
2. Only serum should be used.
3. Avoid grossly hemolytic, lipemic or turbid samples.
4. Preferably use fresh samples. However, specimens can be stored up to 48 hours at 2-8°C, for short duration.
5. For longer storage, specimens can be frozen at -20°C. Thawed samples must be mixed prior to testing.
6. Do not heat inactivate before use.
7. Specimen containing precipitate or particulate matter should be clarified by centrifugation prior to use.
8. Specimen should be free from particulate matter and microbial contamination

PRECAUTIONS

1. Bring all reagents and specimen to room temperature before use.
2. Do not pipette any material by mouth.
3. Do not eat, drink or smoke in the area where testing is done.
4. Use protective clothing and wear gloves when handling samples.
5. Use absorbent sheet to cover the working area.
6. Immediately clean up any spills with sodium hypochlorite.
7. All specimens and standards should be considered potentially infectious and discarded appropriately.
8. Neutralize acid containing waste before adding hypochlorite.
9. Do not use kit after the expiry date.
10. Do not mix components of one kit with another.
11. Always use new tip for each specimen and reagent.
12. Do not allow liquid from one well to mix with other wells.
13. Do not let the strips dry in between the steps.

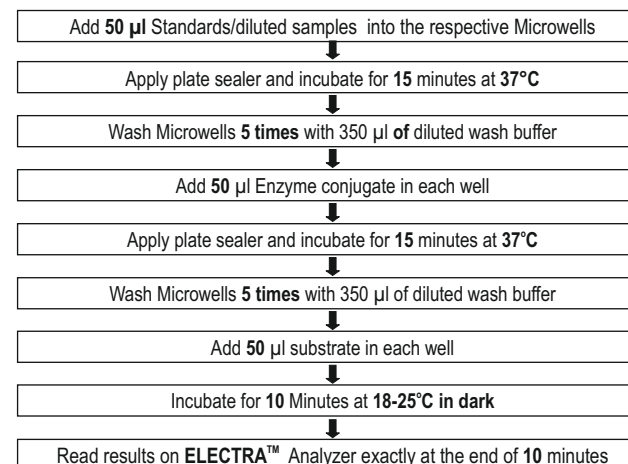
REAGENT PREPARATION

- All reagents should be brought to room temperature (18-25°C) and mixed by gently inverting or swirling prior to use. Do not induce foaming.
- Dilute wash buffer 20 times (for example add 5ml concentrated buffer to 95 ml distilled or deionized water). Mix well before use.
- Prepare a Working Substrate by Mixing Substrate A and Substrate B in equal volume (1:1 ratio) before addition to the micro-wells.

No. of Strips	1	2	3	4	5	6	7	8	9	10	11	12
Substrate- A µl	250	450	650	850	1050	1250	1450	1650	1850	2050	2250	2450
Substrate- B µl	250	450	650	850	1050	1250	1450	1650	1850	2050	2250	2450

TEST PROCEDURE

1. Patient serum should be diluted 51 fold, before use. (i.e mix **10 µl** serum with **500 µl** Sample Diluent).
2. **Important Note: The CA15-3 standards have already been prediluted and are ready for use. Please DO NOT dilute again.**
3. Secure the desired number of coated wells in the holder. Dispense **50 µl** of CA15-3 standards & diluted serums into the appropriate wells. Incubate at 37°C for **15 minutes**.
4. After incubation, empty the microtitre wells and wash the plate 5 times with 350µl of diluted wash buffer. Strike the microtitre plate sharply onto absorbent paper towel to remove all residual droplets.
5. Dispense **50 µl** of enzyme conjugate into each well. Incubate at 37°C for **15 minutes**.
6. Remove the contents and wash the plate as described in step 4 above.
7. Add **50 µl** of working Substrate (A+B) in all the micro-wells. Keep away from direct light while adding the substrate.
8. Cover the **ELECTRA™** microplate and incubate for 10 minutes at room temperature (18-25°C) in dark.
9. Read the **ELECTRA™** micro-plate exactly at 10 minutes in **ELECTRA™ Analyzer**.



CALCULATIONS

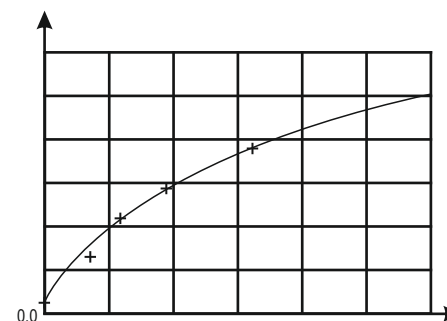
Construct a standard curve by plotting the RLU obtained from each reference standards against its concentration in U/ml on the graph paper, with RLU values on the vertical or Y axis and concentrations on the horizontal or X axis. Use the RLU values for each specimen to determine the corresponding concentration of CA 15-3 in U/ml from the standard curve. Any diluted specimens must be corrected by the appropriate dilution factor.

Example of Standard curve

Results of a typical standard run with RLU's shown in the Y axis against CA 15-3 concentrations shown in the X axis.

Suggest: Use 4-Parameter Standard curve to calculate sample values.

CA 15-3 Values (U/ml)	RLU's
A	1418
B	145815
C	328436
D	765125
E	1731706
F	3139209



This Standard curve is for the purpose of illustration only and should not be used to calculate unknowns. Each user should obtain their own Standard curve and data.

Expected Ranges of values

Healthy women are expected to have CA15-3 values below 35 U/ml. The minimum detectable concentration of CA15-3 in this assay is estimated to be 5 U/ml.