

SPECIMEN COLLECTION

1. Collect blood specimen by venipuncture according to the standard procedure.
2. Only serum should be used.
3. Avoid grossly hemolytic, lipemic or turbid samples.
4. Preferably use fresh samples. However, specimens can be stored up to 48 hours at 2-8°C, for short duration.
5. For longer storage, specimens can be frozen at -20°C. Thawed samples must be mixed prior to testing.
6. Do not heat inactivate before use.
7. Specimen containing precipitate or particulate matter should be clarified by centrifugation prior to use.
8. Specimen should be free from particulate matter and microbial contamination.

PRECAUTIONS

1. Bring all reagents and specimen to room temperature before use.
2. Do not pipette any material by mouth.
3. Do not eat, drink or smoke in the area where testing is done.
4. Use protective clothing and wear gloves when handling samples.
5. Use absorbent sheet to cover the working area.
6. Immediately clean up any spills with sodium hypochlorite.
7. All specimens and standards should be considered potentially infectious and discarded appropriately.
8. Neutralize acid containing waste before adding hypochlorite.
9. Do not use kit after the expiry date.
10. Do not mix components of one kit with another.
11. Always use new tip for each specimen and reagent.
12. Do not allow liquid from one well to mix with other wells.
13. Do not let the strips dry in between the steps.

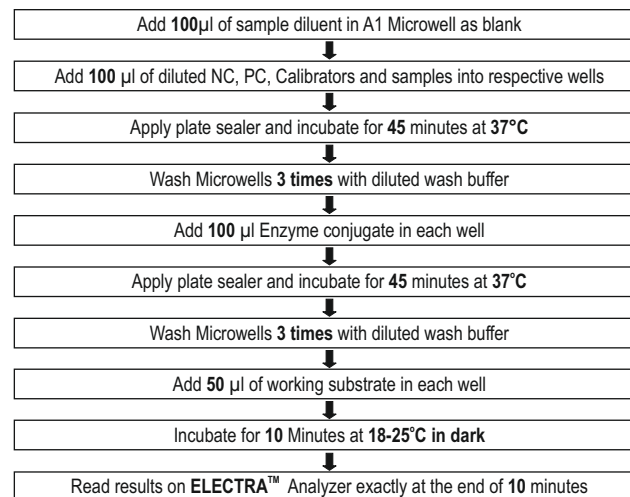
REAGENT PREPARATION

1. All reagents should be brought to room temperature (18-25°C) and mixed by gently inverting or swirling prior to use. Do not induce foaming.
2. Dilute wash buffer 20 times (for example add 5ml concentrated buffer to 95ml of distilled or deionized water).
3. Prepare a working substrate by mixing substrate A and Substrate B in equal volume (1:1 ratio) before addition to the microwells.

No. of Strips	1	2	3	4	5	6	7	8	9	10	11	12
Substrate- A µl	250	450	650	850	1050	1250	1450	1650	1850	2050	2250	2450
Substrate- B µl	250	450	650	850	1050	1250	1450	1650	1850	2050	2250	2450

TEST PROCEDURE

1. Place the desired number of coated strips into the holder.
2. Prepare 1:40 dilutions by adding 5µl of the test samples, negative control, positive control, and calibrators to 200 µl of sample diluent. Mix well.
3. Dispense 100µl of diluted serum samples, negative control, positive control, and calibrator into the appropriate wells. For the reagent blank, dispense 100µl of sample diluent in A1 well position. Tap the holder to remove air bubbles from the liquid and mix well. Incubate for 45 minutes at 37°C.
4. Wash each well three times by filling approximately 350µl diluted wash buffer & blot dry.
5. Dispense 100µl of enzyme conjugate to each well and incubate for 45 minutes 37°C.
6. Wash each well three times by filling approximately 350µl diluted wash buffer & blot dry.
7. Add 50 µl of working Substrate (A+B) in all the micro-wells. Keep away from direct light while adding the substrate.
8. Cover the **ELECTRA™** microplate and incubate for 10 minutes at room temperature (18-25°C) in dark.
9. Read the **ELECTRA™** micro-plate exactly at 10 minutes in **ELECTRA™ Analyzer**. If **ELECTRA™** micro-plate is not read between 10-15 minutes the test results should be considered as invalid.



RUN CRITERIA

The test run may be considered valid provided the following criteria are met:

1. The Rubella IgG Index for Negative and Positive Control should be in the range stated on the labels.

AVIDITY TESTING

Avidity is a measure of antigen to antibody binding. Avidity Test helps in discriminating primary infection from secondary infection. Sometimes it is not sufficient to test for IgM antibodies, as the presence of this class may be due to the persistence of IgM antibodies due to past infection or asymptomatic re-infection without risk for the fetus. For this reason, it is useful to assay the avidity of IgG antibodies. The presence of low avidity is therefore an indication of recent or current infection. The avidity of IgG antibodies can be assayed with this same kit using an additional Buffer called Avidity Buffer (Cat No. 532010096) which is available on request.

For Procedure and Interpretation of results, kindly refer Pack Insert of Avidity Buffer.

CALCULATIONS

Qualitative Determination of Rubella IgG

1. Rubella IgG index value can be calculated by dividing the mean absorbance of NC/PC/Sample by absorbance of Cut-Off calibrator (15 IU/ml).

$$\text{Rubella IgG Index of NC} = \frac{\text{RLU of NC}}{\text{RLU of Cut-Off calibrator}}$$

$$\text{Rubella IgG Index of PC} = \frac{\text{RLU of PC}}{\text{RLU of Cut-Off calibrator}}$$

$$\text{Rubella IgG Index of sample} = \frac{\text{RLU of Sample}}{\text{RLU of Cut-Off calibrator}}$$

Quantitative Determination of Rubella IgG

For a quantitative determination of anti-Rubella IgG levels of specimens in IU/ml unit, RLU of calibrators are plotted on the Y-axis in graph versus their corresponding anti-Toxoplasma IgG concentration 0, 15, 30, and 100 IU/ml on the X-axis. The estimates of levels in patient sera are read off the 4parameter logistic regression curve using their individual RLU values. For example:

For example:

Rubella IgG Values (IU/ml)	RLU's
A	53285
B	6670187
C	8974196
D	9976653

