Chemiluminescence assay

INTERPRETATION OF THE RESULT

IgG Index Value | Result | Interpretation
--- | --- | ---
IgG Index value < 0.90 | Negative | Indicates absence of prior exposure to Rubella (<13 IU/ml)
IgG Index value 0.91-0.99 | Grey zone | Sample should be re-tested, (13-15 IU/ml)
IgG Index value > 1.0 | Positive | Indicates prior exposure to Rubella virus (>15 IU/ml)

PERFORMANCE CHARACTERISTICS

A total of 117 patient samples were used to evaluate specificity and sensitivity of the test. ELECTRA™ Rubella IgG test results were compared to a commercial available licensed ELISA kit:

<table>
<thead>
<tr>
<th>Reference CLIA</th>
<th>ELECTRA™ Rubella IgG CLIA</th>
</tr>
</thead>
<tbody>
<tr>
<td>N</td>
<td>71 (D) 0 0 (B) 71</td>
</tr>
<tr>
<td>E</td>
<td>0 3 0 4</td>
</tr>
<tr>
<td>P</td>
<td>0 (C) 1 41 (A) 42</td>
</tr>
<tr>
<td>Total</td>
<td>71 4 41 117</td>
</tr>
</tbody>
</table>

Sensitivity = 100%
Specificity = 100%

Important Note:
1. This assay is a temperature sensitive assay. The best temperature condition for this assay is 37°C.
2. The wash procedure is critical. Insufficient washing will result in poor precision and falsely elevated RLU readings.
3. It is recommended to use the multiple channel pipettes to avoid time effect. A full plate of 96 wells may be used if automated pipetting is available.
4. Duplication of Calibrators & samples is not mandatory but may provide information on reproducibility & application errors.

LIMITATIONS OF THE ASSAY

1. Lipemic, hemolyzed, icteric or heat inactivated sera may cause erroneous results.
2. Rubella antibody is present in apparently normal subjects in certain populations or geographic groups. A single test is not diagnostic for an active infection. Obtain two specimens at an interval of two weeks and test them at the same time to give more meaningful information.
3. As with other serological assays, the results of these assays should be used in conjunction with information available from clinical evaluation and other diagnostic procedures.

BIBLIOGRAPHY


SYMBOL KEYS

Manufactured by:
Zephyr Biomedicals
A Division of Tulip Diagnostics (P) Ltd.
A Division of Tulip Diagnostics (P) Ltd.
M-46-47, Phase III B, Verna Industrial Estate, Verna, Goa- 403 722. INDIA.
Regd. Office: Gitanjali, Tulip Block, Dr. Antonio De Resg Bagh, Alto Santacruz,™
Bambolim Complex P.O., Goa - 403 202, INDIA.

INTENDED USE

ELECTRA™ Rubella IgG is intended for the Quantitative detection of IgG antibody to rubella virus infection in human serum. For use on ELECTRA™ analyzers.

INTRODUCTION

Rubella is a herpes virus. Generally, rubella is considered a mild adolescence disease; however, a maternal infection could be transmitted through the placenta to the fetus, causing congenital rubella. Congenital rubella may result in chronic cardiac disease, growth retardation, hepatosplenomegaly, malformations, and other severe abnormalities. Children born asymptomatic may develop these abnormalities later in life.

To reduce risk of such severe complications, accurate serological methods must be performed to determine the serologic status of childbearing-aged women. The presence of rubella specific IgG in the bloodstream attests immunity to rubella. An increase in rubella IgG denotes an acute infection and differentiates rubella from other exanthematosis diseases.

PRINCIPLE

ELECTRA™ Rubella IgG CLIA is for use on ELECTRA™ analyzers. ELECTRA™ Rubella IgG CLIA works on the principle of chemiluminescence wherein light is produced by a chemical reaction from a substance as it returns from an electronically excited state to the ground state. When catalyzed by HRP, the oxidation of luminol by hydrogen peroxide produces an electronically excited form of 3-aminophthalate which on relaxation emits light with maximum intensity at λ=425nm.

In this assay Puriﬁed rubella antigen is coated on the surface of microwells. Diluted patient serum is added to wells, and the rubella IgG speciﬁc antibody, if present, binds to the antigen. All unbound materials are washed away. A subsequent incubation with Anti-human IgG agglutinating sera conjugated with horseradish peroxidase binds to the antigen-antibody complex. Excess enzyme conjugate is washed off, and bound enzyme is detected by adding chemiluminescent substrate and Luminescence is measured in RLU. The intensity of the emitting light is directly proportional to the amount of enzymatic activity of the immunocomplex and hence to the amount of rubella IgG antibodies in the test samples.

MATERIALS & COMPONENTS

Materials provided with the test kits:
2. Sample Diluent. Ready to use.
3. Negative Calibrator: 0 IU/mL.
4. Positive Calibrator: 30 IU/mL.
5. Positive Calibrator: 100 IU/mL.
6. Positive Control: Range stated on the label.
7. Positive Control: Range stated on the label.
8. Wash Buffer Concentrate (20X).
10. Cut-oﬀ Calibrator: 15 IU/mL. Rubella G Index = 1.0
11. Substrate A: Chemiluminescent substrate containing enhanced luminol solution. Enzyme conjugate is washed off, and bound enzyme is detected by adding chemiluminescent substrate and Luminescence is measured in RLU. The intensity of the emitting light is directly proportional to the amount of enzymatic activity of the immunocomplex and hence to the amount of rubella IgG antibodies in the test samples.

Materials required but not provided:
1. Precision pipettes: 10-100µl, 20-200µl, 100-1000µl
2. Disposable pipette tips
3. Distilled water
4. Disposable Gloves
5. Automated Washer
6. Avidity Buffer
7. ELECTRA™ Analyzer

STORAGE AND STABILITY

1. ELECTRA™ Rubella IgG kit is stable at 2-8°C up to the expiry date printed on the label.
2. Coated Microwells should be used within one month upon opening the pouch provided that once opened, the pouch must be resealed to protect from moisture. If the color of the desiccant has changed from blue to white at the time of opening the pouch, another coated Microwells pouch should be used.
3. Diluted Wash Buffer is stable up to one week when stored at 2-8°C.
4. Working Substrate (A+B) must be used immediately.
SPECIMEN COLLECTION
1. Collect blood specimen by venipuncture according to the standard procedure.
2. Only serum should be used.
3. Avoid grossly hemolytic, lipemic or turbid samples.
4. Preferably use fresh samples. However, specimens can be stored up to 48 hours at 2-8°C, for short duration.
5. For longer storage, specimens can be frozen at -20°C. Thawed samples must be mixed prior to testing.
6. Do not heat inactivate before use.
7. Specimen containing precipitate or particulate matter should be clarified by centrifugation prior to use.
8. Specimen should be free from particulate matter and microbial contamination.

PRECAUTIONS
1. Bring all reagents and specimen to room temperature before use.
2. Do not pipette any material by mouth.
3. Do not eat, drink or smoke in the area where testing is done.
4. Use absorbent sheet to cover the working area.
5. Immediately clean up any spills with sodium hypochlorite.
6. Use protective clothing and wear gloves when handling samples.
7. Do not eat, drink or smoke in the area where testing is done.
8. Neutralize acid containing waste before adding hypochlorite.
9. Do not use kit after the expiry date.
10. Do not mix components of one kit with another.
11. Always use new tip for each specimen and reagent.
12. Do not allow liquid from one well to mix with other wells.
13. Do not let the strips dry in between the steps.

REAGENT PREPARATION
1. All reagents should be brought to room temperature (18-25°C) and mixed by gently inverting or swirling prior to use. Do not induce foaming.
2. Dilute wash buffer 20 times (for example add 5ml concentrated buffer to 95ml of distilled or deionized water).
3. Prepare a working substrate by mixing substrate A and Substrate B in equal volume (1:1 ratio) before addition to the microwells.
4. For longer storage, specimens can be frozen at -20°C. Thawed samples must be mixed prior to testing.

TEST PROCEDURE
1. Place the desired number of coated strips into the holder.
2. Prepare 1:40 dilutions by adding 5µl of the test samples, negative control, positive control, and calibrators to 200 µl of sample diluent. Mix well.
3. Prepare a working substrate by mixing substrate A and Substrate B in equal volume (1:1 ratio) before addition to the microplate. Mix well. Incubate for 45 minutes at 37°C.
4. Wash each well three times by filling approximately 350µl diluted wash buffer & blot dry.
5. Dispense 100µl of enzyme conjugate to each well and incubate for 45 minutes 37°C.
6. Wash each well three times by filling approximately 350µl diluted wash buffer & blot dry.
7. Add 100µl of diluted NC, PC, Calibrators and samples into respective wells
8. Apply plate sealer and incubate for 45 minutes at 37°C
9. Wash Microwells 3 times with diluted wash buffer
10. Add 100 µl Enzyme conjugate in each well
11. Apply plate sealer and incubate for 45 minutes at 37°C
12. Wash Microwells 3 times with diluted wash buffer
13. Add 50 µl of working substrate in each well
14. Incubate for 10 Minutes at 18-25°C in dark

Calculator
Rubella IgG Index of sample = RLU of Sample / RLU of Cut-Off calibrator
Rubella IgG Index of NC = RLU of NC / RLU of Cut-Off calibrator
Rubella IgG Index of PC = RLU of PC / RLU of Cut-Off calibrator

RUN CRITERIA
The test run may be considered valid provided the following criteria are met:
1. The Rubella IgG Index for Negative and Positive Control should be in the range stated on the labels.

AVIDITY TESTING
Avidity is a measure of antigen to antibody binding. Avidity Test helps in discriminating primary infection from secondary infection. Sometimes it is not sufficient to test for IgM antibodies, as the presence of this class may be due to the persistence of IgM antibodies due to past infection or asymptomatic re-infection without risk for the fetus. For this reason, it is useful to assay the avidity of IgG antibodies. The presence of low avidity is therefore an indication of recent or current infection. The avidity of IgG antibodies can be assayed with this same kit using an additional Buffer called Avidity Buffer (Cat No. 532010098) which is available on request. For Procedure and Interpretation of results, kindly refer Pack Insert of Avidity Buffer.

CALCULATIONS
Qualitative Determination of Rubella IgG
1. Rubella IgG index value can be calculated by dividing the mean absorbance of NC/PC/Sample by absorbance of Cut-Off calibrator (15 IU/ml).
2. Rubella IgG Index of NC = RLU of NC / RLU of Cut-Off calibrator
3. Rubella IgG Index of PC = RLU of PC / RLU of Cut-Off calibrator
4. Rubella IgG Index of sample = RLU of Sample / RLU of Cut-Off calibrator

Quantitative Determination of Rubella IgG
For a quantitative determination of anti-Rubella IgG levels of specimens in IU/ml unit, RLU of calibrators are plotted on the Y-axis in graph versus their corresponding anti-Toxoplasma IgG concentration 0, 15, 30, and 100 IU/ml on the X-axis. The estimates of levels in patient sera are read off the 4parameter logistic regression curve using their individual RLU values. For example:

For example:

<table>
<thead>
<tr>
<th>Rubella IgG Values (IU/ml)</th>
<th>RLU's</th>
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</thead>
<tbody>
<tr>
<td>A</td>
<td>53285</td>
</tr>
<tr>
<td>B</td>
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<tr>
<td>C</td>
<td>8974196</td>
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<tr>
<td>D</td>
<td>9976653</td>
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Graph showing the relationship between RLU values and Rubella IgG levels.